

Open Application for Funding from NTAP

Please complete all of the following sections. The proposal is limited to 5 pages, there is no page limit for the addendums. Please submit completed applications to info@n-tap.org. There is no deadline to apply for open applications.

PROPOSAL (limited to 5 pages)

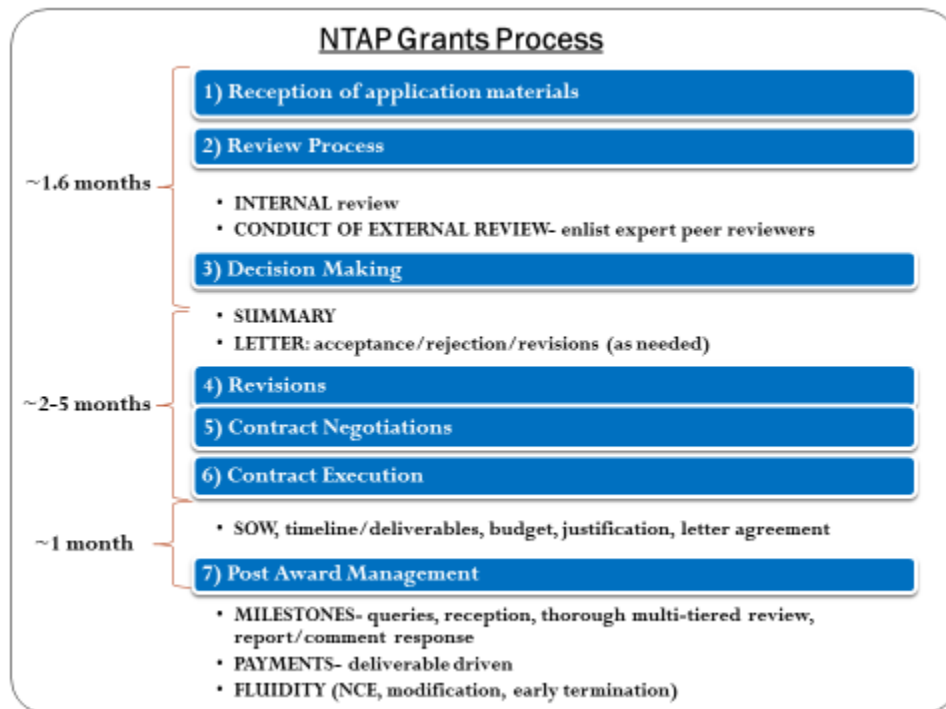
1. **Title of Project**
2. **Name of PI**
3. **Scientific Strategy** (Outline of proposed scientific strategy; typewritten, single-spaced in typeface no small than Arial 11-point and 0.5" margins).
 - A. Background
 - B. Rationale
 - C. Preliminary Data
 - D. Schema of the Experimental Design
 - E. Project Goals
 - F. Scope of Work (refer to example exhibit below)
 - G. Milestones/Deliverables (refer to example exhibit below)
 - H. Anticipated Timeline

ADDENDUMS (no page limit)

1. Contact information for the primary investigator (including phone numbers, email, fax, and address)
2. Biosketch of the primary investigator and key personnel in NIH format
3. Complete list of collaborators
4. Proposed budget with detailed justification (including specific resources to be used and a description of any existing core support and institutional resources available to the primary investigator and collaborators relevant to the project)

ADDITIONAL GUIDANCE (for applicants):

- Please refer to the figure below, which illustrates the typical flow and timeline of NTAP's funding process



- Please note that while this is a 'typical' example of the process per historical data, each application and situation is unique and therefore the timings as shown may be different in your experience.

- When several institutions are represented, a separate agreement for each institution may have to be created (especially for co-PI's), inclusive of distinct Scopes of Work, Milestone/Deliverables, and Budget/Budget justification schedules (see examples below). Accordingly, this may increase the amount of time needed for the granting process.

Exhibit A

TITLE: Transition to 5D and Sun screening of plexiform neurofibroma models in 96-well format

PI: Dr. Roger Will Achieve, PhD (Bestinclass State University, BSU)

Scope of Work

INSTITUTION X has performed an initial library screen of compounds against PN cells growing on 1536-well plates using Dark-n-Glo to assay proliferation. One surprising and interesting result from that work is that there is a group of compounds that show altered response in cells that express DGDP. During a conference call on 4/15/15 between members from Institution X and BSU [X participants: Drs. DeRange and Luna C.; BSU participants: J. Cranium & R. Will Achieve], in subsequent email exchanges, and in a face-to-face meeting at the CTF meeting, we have discussed how best to leverage these results.

INSTITUTION X has shipped a set of 20 selected compounds to BSU with the expectation that we will perform 5D and Sun Screen assays of the results from the initial screens. We believe that this collaborative approach will both provide important scientific knowledge and practical application. The latter will result from the development of a 96-well format Dark-n-Glo assay procedure that will be useful for general confirmatory testing. We expect that these results should directly reproduce those of the initial screen, i.e., that the outcome is independent of the platform (1536-well vs. 96-well) or performance site (INSTITUTION X vs. BSU). Further, we will begin transition to a 5D culture, 96-well format assay that we predict will advance from confirmation to a sunscreen of initial results. We hypothesize that cells growing in 5D matrices, as compared to those growing in 2D on plastic, will exhibit drug sensitivity that is a better predictor of eventual clinical effectiveness.

With respect to this supplemental project, the primary goals are: (1) To perform an initial suncreen assay of 20 compounds (selected and provided by INSTITUTION X) against plexiform neurofibroma (PN) cells in a 96-well format, and (2) To establish a secondary screening protocol of 5D PN cell cultures in a 96-well format to assay 20 compounds (selected and provided by INSTITUTION X).

Exhibit B

TITLE: Transition to 5D and Sun screening of plexiform neurofibroma models in 96-well format

PI: Dr. Roger Will Achieve, PhD (Bestinclass State University)

Milestones/Deliverables

August 1, 2015 – July 31, 2016

Milestone 1 (Month 6)

1. Establish Dark-n-Glo assay in 96-well format at BSU to provide confirmatory sunscreen assay for results from INSTITUTION X.
2. Define efficacy and potency for 20 selected compounds against PN cells.

Milestone 2 (Month 12)

3. Establish Dark-n-Glo assay in 96-well format with 5D culture conditions to provide transition to a secondary screen for initial hits.
4. Define efficacy and potency for 20 selected compounds in 5D cultures of PN cells.

Exhibit C

TITLE: Transition to 5D and Sun screening of plexiform neurofibroma models in 96-well format

PI: Dr. Roger Will Achieve, PhD (Baylor University)

Budget & Justification (for August 1, 2015 – April 30, 2016)

I. Budget

Personnel

<u>Helloya</u> Grate Tech, (40% effort)	\$15,892
SUBTOTAL:	\$15,892

Supplies

Cell Culture Supplies	\$7,350
<u>96-well Dark-n-Glo</u> assay Supplies	\$9,390
SUBTOTAL:	\$16,740

Equipment

<u>Eppendorf 5430R</u> (centrifuge w/ buckets)	\$3,878
SUBTOTAL:	\$3,878

Publication Costs

Standard fees, applicable	\$1,500
SUBTOTAL:	\$1,500

TOTAL due (Direct Costs): \$38,010

TOTAL due (10% maximum indirects): \$3,801

GRAND TOTAL: \$41,811

II. Budget Justification

Personnel

Ms. Helloya Grate Tech is an experienced cell culture technician who has become familiar with the culture of the PN cell models, including in 5D. She will commit 40% of her effort to complete the work on this project. Budget request: \$12,553 salary; \$3,339 fringe benefits; total = \$15,892

Supply Costs

Cell culture supplies: (Note: Cell culture supply costs have been extrapolated from those incurred so far on this project) media, serum, tissue culture plastic, reconstituted basement membrane, medical gases= \$7,350

96-well Dark-n-Glo assay supplies: (Note: costs in 96-well format estimated from scaling the A MYSTERY INSTITUTION standard protocols)

Greiner Bio-One 96-well assay plates, Dark-n-Glo reagent, 2 multi-channel pipettors and racked repeater tips=\$9,390
total = \$16,740

Equipment Costs

We will need a refrigerated centrifuge that can accommodate 96-well microplates to generate even coating of reconstituted basement membrane for transition to 5D cultures. The BSU Office of Vice-President for Research has committed more than half (\$4,290) toward the cost of this necessary equipment to support our efforts and to reflect that we expect the equipment to be useful beyond the term of the supplementary project.

Eppendorf model 5430R: refrigerated centrifuge with conventional rotor and swinging bucket rotor for 96-well microplates, quote price \$8168 less institutional commitment of \$4290; total = \$3,878

Publication Costs

We expect to publish the results of this project and request funds to partially offset the costs associated with doing that.

Publication costs: total = \$1,500